PHOSPHATASE TEST - CHARM® FAST ALKALINE PHOSPHATASE TEST USING CHARM NOVALUM® AND NOVALUM II X® IMS #28c

[Unless otherwise stated all tolerances are ±5%]

SAMPLES

1.	Laboratory Requirements (see Cultural Procedures [CP], items 34 & 35) [See current version of M-a-98 to determine if this test method has been approved for use on the specific dairy product being tested]				
	a.	Product Groups/Descriptions			
		Fluid white milks – including skim through whole fat milk			
		Unflavored liquid dairy products – including half and half, cream, light cream, whipping cream (products that can be accurately pipetted)			
		3. Flavored liquid dairy products (Liquid products that can be accurately pipetted, containing flavor additives and/or thickening agents including flavored milk, and etc.)			
		APPARATUS			
2.	. CP, items 1-33 (as necessary)				
	a.	Unless otherwise stated, "shake vigorously" refers to standard microbiological mixing, i.e., 25 times in a 1 foot movement in 7 sec or vortex for 10 sec at maximum setting (subsamples/controls in an appropriate container for vortexing)			
3.	Pipe	ettors and Pipets			
	a.	Fixed volume or electronic, 100 μL			
	b.	Calibration checked as specified in CP item 6.e; maintain records			
	C.	Disposable, 10 mL (ASTM) pipet with 0.1 mL graduations			
4.	Micı	rotube Adapter for NovaLUM/NovaLUM II X			
5.	Nov	vaLUM/NovaLUM II X Analyzer			
	a.	Operating instructions available			
		Channels configured for Fast Alkaline Phosphatase (FAP) assay for appropriate definitions			
		a. FAP MILK – 45 sec time			

			b. FAP CREAM – 90 sec time			
			c. FAP CHOC – 90 sec time			
		2.	Thermoprobe connected with NovaLUM (positioned upright in stand) or NovaLUM II X			
			 a. Probe measuring ambient room temperature (must be between 18-24°C to run the test) 			
		3.	Microtube adapter for Luminometer/Luminator/NovaLUM/NovaLUM II X			
6.	13 x		ath, Circulating, 34±1°C and 63±1°C (or 66±1°C if fat > 10%), or Test Tube Dry Well Heater Blocks Acceptable (Confirmation			
7.			ge – Charm II Heraeus® (3,400 RPM), Minifuge, or Equivalent ,000 g)			
8.	Vor	tex N	lixer			
9.	Han	dling	uge – Charm II Heraeus® (3,400 RPM), Minifuge, or Equivalent 2,000 g) Mixer ng and Storage t contains Reagent FAP Vials and Calibrator Tablets t: Lot #: Exp Date:/			
	a.	Kit c	contains Reagent FAP Vials and Calibrator Tablets			
		Kit:	Lot #: Exp Date:/			
		Cali	ibrator Lot #: Exp Date:/			
	b.	Rea	agents stored at 0.0-4.5°C until expiration date			
		1.	FAP vials may be stored at room temperature. If stored at room temperature, laboratory expiration date is 3 weeks from first date of room temperature storage. FAP vials must be at 18-24°C at time of use			
			CONTROLS			
10.	Neg	ative	e Calibrator/Control			
	a.	neg	duct group. Prepare at least 20 mL of negative sample for use as a attive calibrator/control and to rehydrate 350mU/L positive brator/control			
		1.	Fluid white milk – heat a sample of product (highest fat content) to 95±1°C for 1 min with stirring			
		2.	All flavored liquid dairy products can be tested on the FAP CHOC channel by heating a chocolate sample (highest fat content) to 95±1°C for 1 min with stirring			
			a. Cool rapidly in an ice bath and hold at 0.0-4.5°C			

			b. Centrifuge for 3 min and decant supernatant	
		3.	All unflavored liquid dairy products can be tested on the FAP CREAM channel by heating pasteurized light cream to 95±1°C for 1 min with stirring	
		4.	Note: if product precipitates during negative sample preparation, e.g. sheep milk, heating sample to 63°C for 45 min is acceptable. If using 13 x 100 test tube dry well heater block at 95°C, it takes 10 min to heat product to 95°C; once at temperature, time for 1 min (Use TC)	
	b.	Coo	ol rapidly in an ice bath and hold at 0.0-4.5°C	
	C.		re at 0.0-4.5°C, the Negative Control/Sample may be used for up to	
	d.	proc	within 24 hours, aliquot 1 mL quantities into small tubes (see 5.a.1 for duct definitions) seal and freeze at -15°C or colder in a non-frost-free freezer an insulated foam container in a frost-free freezer, use within 2 months	
		Lab	Prep. Date: Lab Exp. Date:	
11.	Pos	itive	350 mU/L Calibrator/Control	
	a.	Prep	pare Positive Calibrator/Control	
		1.	Rehydrate a calibrator tablet with 100 µL MS or DI water, mix to disperse tablet, wait 1 min and mix again	
		2.	Add 2.5 mL of Negative Calibrator/Control (item 10) to dissolve calibrator Tablet	
		3.	Shake vigorously or vortex and let settle 10 min at 0.0-4.5°C for re-suspension	
		4.	Shake vigorously or vortex again and use for test	
	b.	Pos	sitive calibrator/control held at 0.0-4.5°C may be used for 48 hours	
			CALIBRATION	
12.	With Ada		ch New Kit Lot # Calibrate Analyzer and Replace Microtube	
	a.		pare Negative Calibrator/Control and Positive Calibrator/Control,	

	 Select appropriate channel for calibration and follow prompts. Note: Previously calibrated channels will list a selection menu, select 'calibrate'; follow prompts 			
		1.	Test a negative calibrator/control, item 14.c	
		2.	Test a positive calibrator/control, item 14.c	
		3.	Instrument will make internal adjustments	
		4.	Test another negative calibrator/control, item 14.c	
		5.	Test another positive calibrator/control, item 14.c	
		6.	If performance of negative (<15) and positive is in range (320-400), instrument will prompt calibration successful. If performance out of range, instrument will recalculate settings and prompt to perform another positive and negative calibrator/control	
		7.	Repeat steps 4-6. If out of range NovaLUM/NovaLUM II X will prompt a re-calibration, step 1	
			DAY OF USE PERFORMANCE CHECKS	
13.	Each Day of Use, Test a Negative Control/Sample (item 10) and Positive Control (item 11), For at Least One Product			
	a.	Verify FAP vial stored at room temperature. For NovaLUM, select 'programmed plans', select appropriate FAP channel and select menu 3 'Control Check'. For NovaLUM II X, select FAP from home screen, select appropriate pre-programmed FAP channel, then select 'Control Check' and follow prompts		
		1.	Test positive calibrator/control, item 14.c. Positive Control valid, 247-453 mU/L	
		2.	Test negative calibrator/control, item 14.c. Negative Control valid or less than or equal to 15 mU/L	
	b.	Peri	iodic rotation of channels is recommended when multiple channels are used	
			TEST PROCEDURE	
14.	Pro	cedu	re [Samples kept at 0.0-4.5°C throughout testing]	
	a.	Pre	pare sample	
		1.	Mix retail milk samples by inverting container top to bottom, then bottom to top (a complete half circle or 180 degrees) without pausing, 25 times; use within 3 min	

	2.	25 t	negative control or subsamples of retail containers by shaking imes in 7 sec with a 1 ft movement or vortex at least 10 sec at kimum setting; use within 3 min (sample(s)/control(s) must be ppropriate container to allow the use of vortexing)			
	3.	For	flavored dairy products (not including controls, items 10 & 11)			
		a.	Add 1 mL of sample into an appropriate tube or vial (NOT FAP vial)			
		b.	Centrifuge for 3 min			
		C.	Use liquid phase in item 14.c			
b.	Veri	fy FA	AP vial stored at room temperature			
	1.	Pier	ce foil top with clean pipet tip			
C.	(iter	Dispense 100 µL of the prepared sample (item 14.a) or mixed controls tems 10 & 11) into the FAP vial liquid and then immediately press enter n NovaLUM or press the 'Run Test" icon on NovaLUM II X				
	1.	Follo setti	ow prompt and vortex FAP vial with sample for 5 sec at maximum ing			
	2.	fully	ow prompt and attach microtube adapter to threaded side of vial. Then insert vial into NovaLUM/NovaLum II X chamber. This step must be apleted while screen is flashing (30 sec)			
d.	At the end of pre-programmed time, the screen will stop flashing and count the sample. The mU/L phosphatase level will be displayed on screen. For NovaLUM, press OK to print and prepare for next sample. For NovaLUM II X result prints automatically, press the right arrow icon then select 'Run Test' to prepare for the next sample					
e.		Samples with ≥ 350 mU/L of ALP activity are suspect positive and must be confirmed (item 15)				
			CONFIRMATION			
Pos	itive	Conf	firmation			
a.			lab pasteurized negative control and positive control made of edairy product			
b.			trols to verify they are in range. If out of range, recalibrate channel and rols to verify calibration			
c.	Rete	est su	uspect positive sample			
d.			with ≥ 350 mU/L of ALP activity are suspect positive and must			

15.

16.	Mic	Microbial Phosphatase/Heat Stable Phosphatase				
	a.		at 1.0 mL of suspect sample at 63±1°C for 30 min, stirring or mixing ery 10 min (Use TC)			
		1.	If fat content is >10%, heat at 66±1°C for 30 min			
	b.	Coc	ol sample rapidly to 0.0-4.5°C in an ice bath			
	C.	Tes	et positive and negative controls (item 15.a) following item 14			
	d.		st heated sample and unheated sample (original sample) following ———————————————————————————————————			
	e.	e. Interpretation				
		1.	Controls test as specified in item 13			
		2.	If heated and unheated samples have equal activity (-30%, mU/L or RLU), the sample is regarded Not Found for residual phosphatase, the activity originally measured is microbial			
		3.	If the heated sample is more than 30% below unheated sample (mU/L or RLU), the sample contains milk phosphatase activity, either residual or reactivated			
17.	Rea	activated Phosphatase				
	a. Magnesium acetate solution commercially available					
	b.	Or,	Or, prepared in laboratory			
		1.	Dissolve 35.4 g of Mg acetate tetra-hydrate, Mg (C ₂ H ₃ O ₂) ₂ •4H ₂ 0 in 25 mL MS or DI water, warming slightly to aid dissolution			
		2.	Pour solution into 100 mL volumetric flask, rinse original container several times and add rinse to flask			
		3.	After cooling to room temperature, make up to 100 mL (stable for 1 year at 0.0-4.5°C)			
	C.	Pro	cedure			
		1.	Label two 13 x 100 test tubes for appropriate for volume as "Blank" and "Test"			
		2.	Add a 5.0 mL aliquot of sample (unheated, original sample to each test tube			
		3.	Add 0.1 mL DI or MS water to the sample labeled "Blank", and 0.1 mL Mg acetate solution to the sample labeled "Test"			
		4.	Cap tubes, mix and heat both aliquots for 1 hour at 34±1°C (Use TC)			

		5.	in an ice bath	
		6.	Dilute 1 mL of sample containing Mg acetate (Test) with 5 mL (1:6 dilution) of negative control product (item 15.a) and mix, label tube as "Diluted Test"	
		7.	Test undiluted sample containing no Mg acetate (Blank) and diluted sample containing Mg acetate (Diluted Test) for phosphatase activity following item 14	
	d.	Inte	rpretation	
		1.	If the diluted aliquot containing Mg acetate (Diluted Test) has equal (±30%) or greater phosphatase activity than the undiluted aliquot containing no Mg acetate (Blank), the sample is regarded as Not Found for residual phosphatase, and the phosphatase originally measured is of reactivated origin	
			Diluted w/Mg (Test) ≥ Undiluted (Blank) = Reactivated	
		2.	If the diluted aliquot (Diluted Test) contains less (30% below or less) activity than the undiluted aliquot (Blank), the sample is considered Positive for residual phosphatase	
			Diluted w/Mg (Test) < Undiluted (Blank) = Residual	
		3.	A false-positive for residual phosphatase may also be obtained if a reactivatable sample has been allowed to stand at elevated temperatures (20°C) for periods of 1 hour or more before testing (SPC < 20,000/mL)	
			RECORDING, INTERPRETATION, AND REPORTING	
18.	Rec	ordir	ng and Interpretation	
	a.	Rec	ord Values	
	b.	o. Interpret		
		1.	If value obtained is <44 mU/L for fluid white milk or <88 mU/L for flavored/unflavored, the sample is Not Detected	
		2.	If value obtained is ≥350 mU/L or mU/kg, the sample is actionable	
19.	Rep	ort		
	a.	Not	Found for residual phosphatase if:	
		1.	<350 mU/L	

	2.	≥35	0 mU/L but:	
		a.	Meets reactivated phosphatase criteria (item 17.d.1)	
		b.	Meets microbial phosphatase criteria (item 16.e.2)	
		C.	Documentations showing the products was treated in such a way that reactivated phosphatase may be present	
b.	Pos	itive	for residual phosphatase if:	
	1.	≥35	0 mU/L or mU/g and:	
		a.	Meets residual phosphatase criteria (item 17.d.2)	
		b.	No microbial phosphatase present (item 16.e.3)	
		C.	No documentation to show the product could have become Reactivated	